Enzymax, LLC

Product Information

Product Name: EZ DNA Gel Extraction Kit
Catalog No.: EZC203
Size: 50 preps

Related products:	Catalog No.
DNA mini spin columns (30-50µl vol.)	EZC101
DNA Tini spin columns (5-20µl vol.)	EZC106
DNA Gel extraction kit with Tini column (50)	604
PCR Clean-up kit (remove<40mer) (50)	EZC 201

Features:

- Easy and rapid with 20 min procedure using spin column
- High DNA recovery
-Extract and purify DNA fragments (70 bp-12 Kb) from standard or low-melting agarose gel in TAE or TBE buffer

Kit Contents:

Components	*EZ DNA Gel extraction 50 Preps* (cat# EZC203)
Gel Extraction Buffer	25 ml
5xWash Buffer*	6 ml
Mini spin column with collection tube**	50

***Add 100% ethanol before use:** add 24 ml 100% ethanol to6 ml 5xWash buffer.
**Mini Spin columns can be order separately for leftover solutions (cat#EZC101, $36 for 100 columns)

Caution:

Gel Extracion Buffer contains chaotropic salt. Please use proper safety precautions and always wear gloves when handling the reagent. Avoid contact with skin, eyes or clothing. In case of accidental spill or contact, wash thoroughly with water, seek medical advice if necessary.

Protocols:

This kit is designed for DNA fragment recovery from agarose gel using Mini spin columns (Cat#EZC101). If you have small amount of DNA sample on the gel, you can use Tini Spin Columns (EZC106) with same protocol but cut down all the solution to1/2 to 1/3. The elution volume for Tini spin column can be as low as 5µl.

1. Excise the DNA fragment from the gel with a clean scalpel, weight and transfer it to a clean tube.
2. Add 400µl **Gel Extracion Buffer** into each 100mg of gel slice.
3. Incubate at 60°C for about 10 min till the gel slice is completely dissolved. Increase the temperature to 85°C, incubation time, or add more extraction buffer if the gel concentration is more than 2%. **Note:** If the color of the mixture turns a blue or purple color, adjust pH by adding a small volume of 3M Sodium acetate (pH 5.0).
3. Load the sample mixture onto the **Mini Spin Column** and spin in a microcentrifuge for 1 min at full speed (about 10,000 rpm). Do not load more than 700 µl of sample on the Mini spin column at one time. Discard the flow through and load more sample mixture if needed.
4. Wash the column by adding 500 µl of **Wash Buffer** (**ethanol added**) and centrifuge for 1 min.
5. Wash once by adding 500 µl of 80% ethanol and centrifuge for 1 min.
6. Discard flow through and place the column back in the same tube. Cut off the cap on the column (this will help to remove ethanol completely) and spin for 2 min. **Note:** this step is important since the residual ethanol may affect downstream applications)
7. Place the column in a clean 1.5 ml micro-centrifuge tube.
8. Add 30 µl or more **distilled water** or **TE** (10mM Tris/HCl, 1mM EDTA, pH8.0) (preheated at 65°C for better yield) to the **center** of the column and leave at room temperature for 5 min. Spin the column for 1 min to elute DNA from the column.

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Enzymax LLC, 870 Corporate Dr. Suite 201, Lexington, KY 40503
Tel. (859) 219-8482; Fax. (859) 219-0653; Web: www.enzymax.net